

Optimization and scale-up of fengycins production by fermentation in chemically defined or co-products based media

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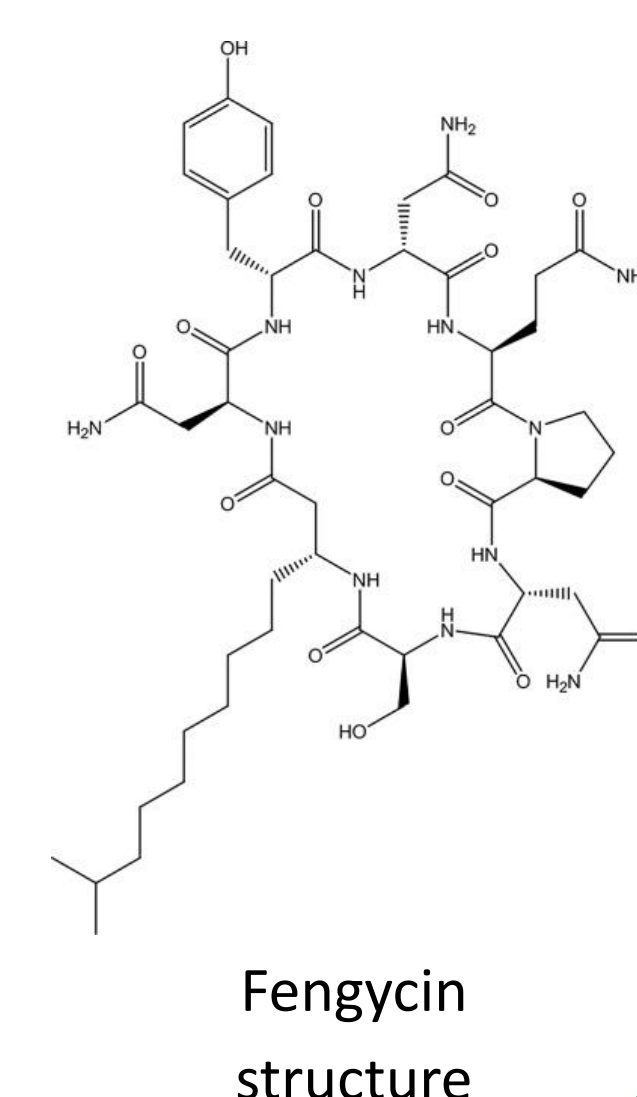
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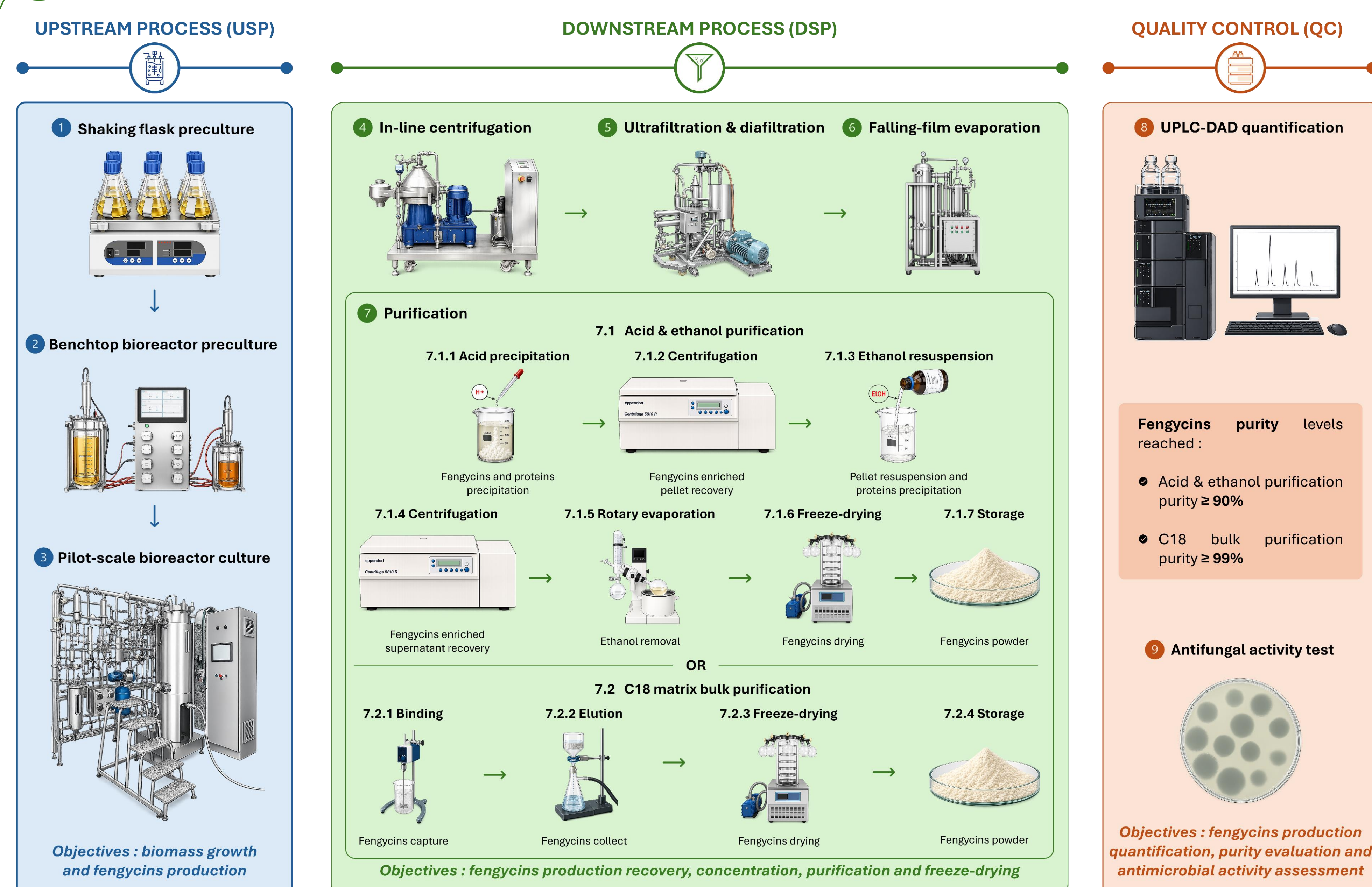
1 RESEARCH CONTEXT

Among all lipopeptides produced by *Bacillus* strains, we are interested in **fengycins**. These cyclic lipopeptides naturally produced by *Bacillus subtilis* are recognized for their strong antifungal, antibiofilm and surfactant properties. These properties lead to many interesting applications in the environmental, agricultural or health sectors.

However, due to their current high production costs, fengycins are in most applications not able to challenge the chemically synthesized counterparts economically. To improve their production yields and make fengycins cost-effective, we implemented an integrated strain engineering strategy. A first production strain (mono- and hyper-producer of fengycins) has been generated through UV-induced mutagenesis (Lipomme-bio project – See poster 1.12). Through CRISPR-Cas9 engineering (SURFs UP project), we then obtained a non-sporulating *B. subtilis* strain producing exclusively fengycins up to 1 g/L. This strain was subsequently used to **develop an efficient and scalable production process**.



2 PRODUCTION PROCESS

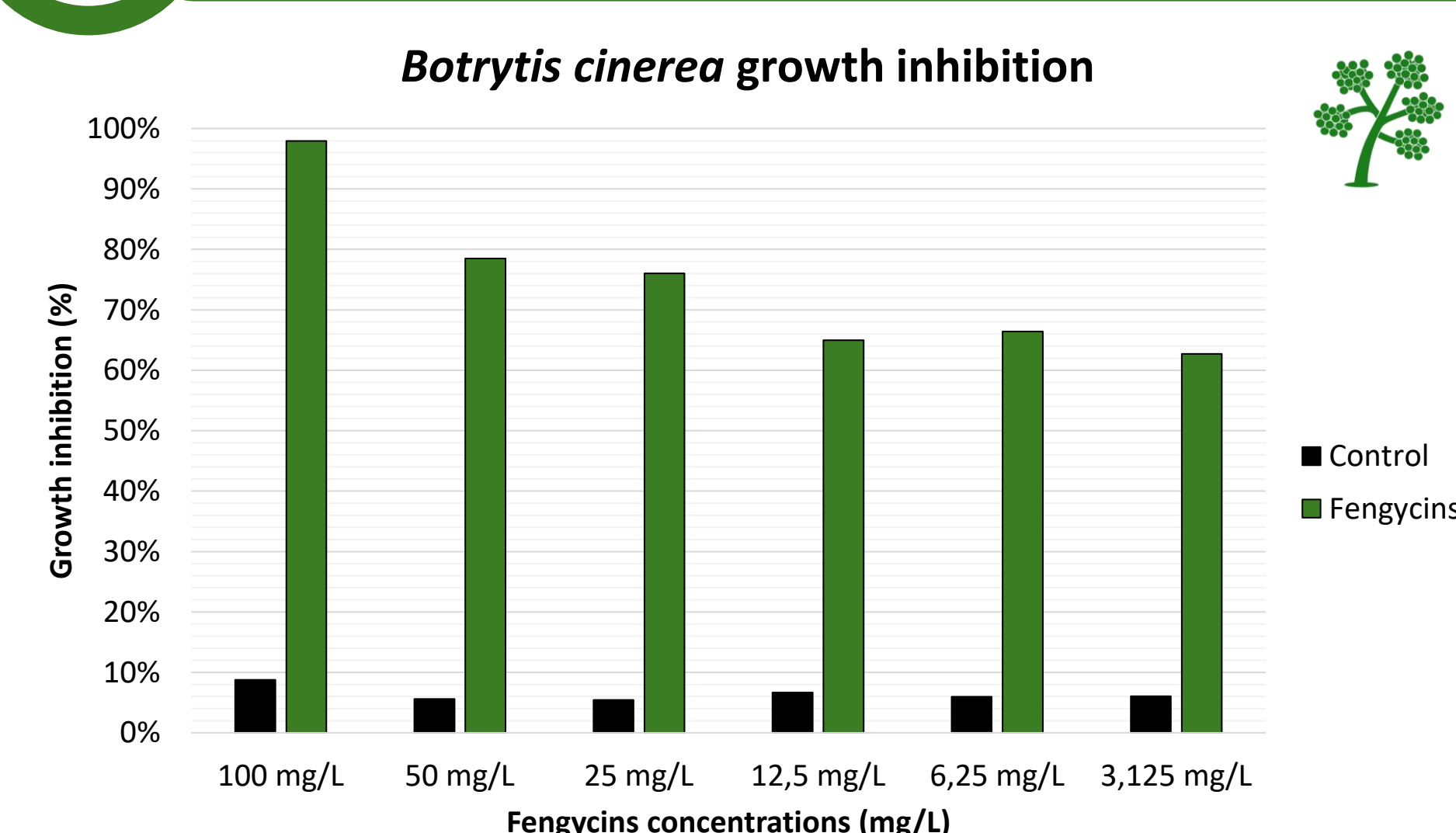


The **upstream process parameters** were first **optimized at bench scale** to identify the most efficient production conditions of our production strain in Landy2X medium.

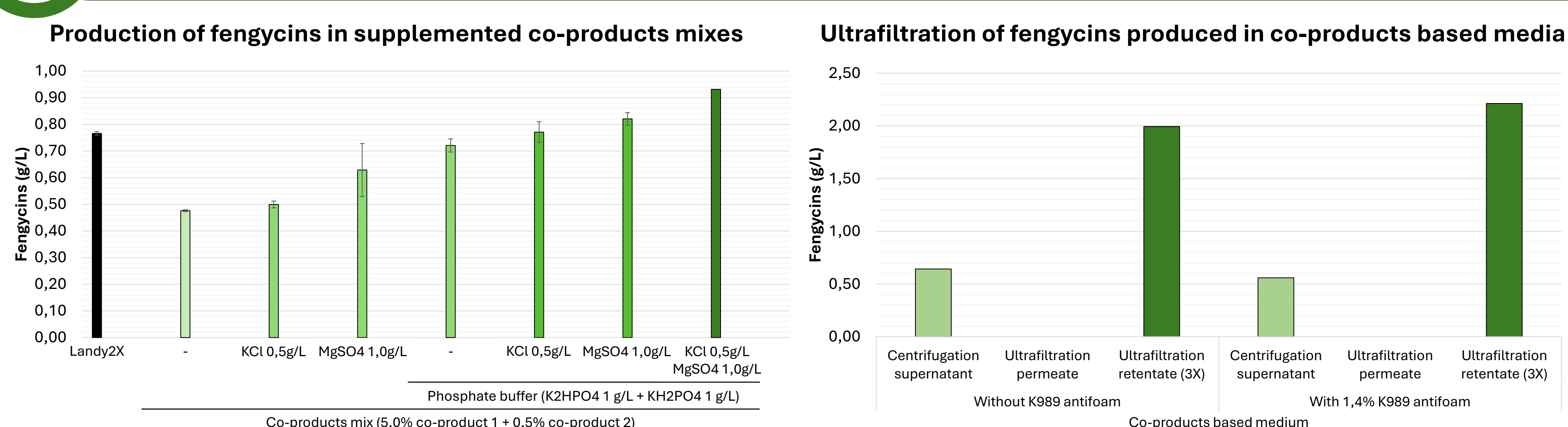
The **optimized upstream process** parameters were then successfully transferred to a 200L pilot-scale bioreactor, enabling the production of approximately **100 grams of fengycins** from 150L of Landy2X medium, demonstrating **strong robustness and scalability** under pre-industrial conditions.

The **developed downstream process** consists of a sequential workflow including clarification by centrifugation, concentration and purification by ultrafiltration and diafiltration (10kDa membrane), and concentration by evaporation steps. **Two different purification processes** yielding **high-purity fengycins** were implemented to further enrich the product, confirming the **efficiency and consistency** of the overall integrated process. Finally, the activity of fengycins is assessed through their antifungal activity.

3 ANTIFUNGAL ACTIVITY



4 PRODUCTION MEDIA



5 RESEARCH HIGHLIGHTS

- ✓ Generation of a non-sporulating *B. subtilis* strain, mono- and hyper-producer of fengycins.
- ✓ Development of an integrated USP–DSP process yielding ± 100 g of fengycins per batch with $\geq 90\%$ purity.
- ✓ Confirmation of the antifungal activity of fengycins against *B. cinerea*.
- ✓ Development of a co-products based medium reducing medium cost by ± 20 folds while maintaining high fengycins production levels.

6 PERSPECTIVES

- 🔍 Optimize fengycins production parameters in the developed co-products based medium.
- 🔍 Scale-up fengycins production process (USP & DSP) in this medium.



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